

Lethality Treatment Determination

Calculating Thermal Inactivation of Pathogens

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AMIFoundation

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Process lethality spreadsheet

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Instructions for Using the AMI Process Lethality Determination Spreadsheet



Objective

The purpose of the process lethality determination model is to provide processors with a science-based validation tool that can be used to demonstrate the effectiveness of a specific heat process to destroy a microorganism of concern. Specifically, the interactive model allows the user to input actual in-process data from a given cook cycle and determine if the process achieves the required log reduction for the microorganism of concern. The goal is to define or map the heating and cooling profile of the product by observing the temperature characteristics of the product during heating and cooling. Microbial destruction may occur during a significant portion of the heating and cooling process, not just at the minimum internal temperature.

American Meat Institute

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How to use the AMI Spreadsheet

1. Select a D, T_{ref} and z value for your product.

- Decide on the pathogen
 - For multiple pathogens use the most thermally resistant (largest D close to finished internal temp)
- Use a published value for a product that is closest in type and composition to your situation
- Hierarchy of importance
 1. Cured vs uncured
 2. Fat level
 3. Species
 4. Whole vs ground vs emulsified

2. Enter data from values recorded in the process

- Use core temperature or the coldest point in the product
- 20 points is sufficient IF enough of the data is in the region above 120F (at least 10 data points)
- May or may not use the cooling part of the process

3. Samples are provided by AMI

Food Research Institute, University of Wisconsin- Madison, July 2012

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Definitions:

D-value: The time (in minutes) at an associated T_{ref} required to kill 90% of the selected microorganism; a one log reduction.

z-value: The number of degrees F to change the D-value by a factor of ten.

F-value: The process lethality. The equivalent time of heating at a reference temperature. Total lethality will be the final computed cumulative F value.

TABLE 1: EXAMPLE - Lethality Data from Literature

Organism	Product	Microbial Heat Tolerance		
		T _{ref} (°F)	z (°F)	D (min)
<i>Salmonella</i>	Meat Patty (Scott and Weddig, 1998)	150	10	0.172
	Gr. Beef (25% fat) (Juneja, 2003)	140	14.5	4.72
<i>E. coli</i> O157:H7	Lean Gr. Beef (2% fat) (Line et al., 1991)	145	8.3	0.30
	Gr. Beef (25% fat) (Juneja, 2003)	140	11.4	3.39
	Lean Gr. Turkey (Juneja and Marmer, 1999)	149	11.7	0.29
	Lean Gr. Lamb (Juneja and Marmer, 1999)	149	12.4	0.38
	Lean Gr. Pork (Juneja and Marmer, 1999)	149	11.7	0.30
<i>Listeria monocytogenes</i>	Lean Gr. Beef (2% fat) (Fain, et al., 1991)	145	9.3	0.6
	Gr. Beef (25% fat) (Juneja, 2003)	140	12.0	4.18
	Hot Dog Batter (30% fat) (Mazzotta and Gombas, 2001)	144	10.8	3.3

Note: This model is a tool for calculating F-values. To ensure correct results, the proper z, T-ref, and D-values for each product and organism must be used.

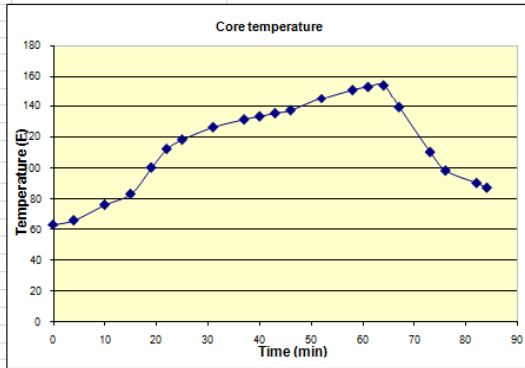
Other values can be found in the handout references

- But be careful on converting units for z values (don't incorporate the 32° offset between F and C)

PROCESS LETHALITY DETERMINATION

Date: August 25, 2010
 Organism: *L. monocytogenes*
 Product name: Hot Dogs

User Must:
 1. Identify organism and product of concern
 2. Provide at least 20 time/temp data points

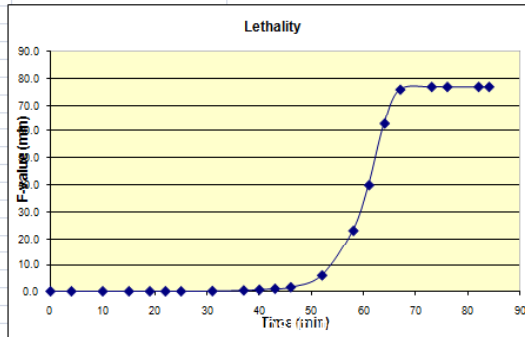


T ref= 144 °F
 z = 10.8 °F
 D = 3.3 min

Log Reduction of Process
 23.30

Time (min)	Core Temp (°F)	F-value (min)
0	63	0.000
4	66	0.000
10	76	0.000
15	83	0.000
19	100	0.000
22	112	0.002
25	118	0.009
31	126	0.086
37	131	0.338
40	133	0.576
43	135	0.940
46	137	1.497
52	145	5.884
58	151	22.941
61	153	39.832
64	154	62.699
67	139	75.864
73	110	76.899
76	98	76.900
82	90	76.900
84	87	76.900

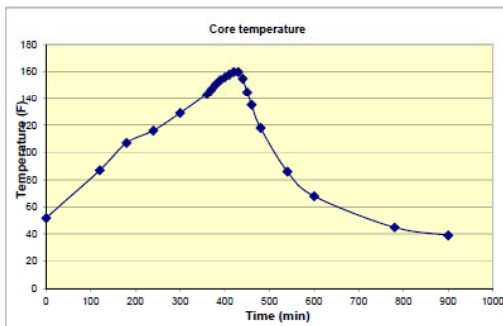
Use the average or low value of internal temperature



PROCESS LETHALITY DETERMINATION

28-Sep-11
 Salmonella
 Ham of 9/23

User Must:
 1. Identify organism and product of concern
 2. Provide at least 20 time/temp data points



T ref= 150 °F
 z = 10 °F
 D = 0.172 min

Log Reduction of Process
 2258.13

Time (min)	Core Temp (°F)	F-value (min)
0	52	0.000
120	87	0.000
180	107	0.002
240	116	0.015
300	129	0.285
360	143	6.489
370	147	9.993
380	151	18.793
390	154	37.648
400	156	70.112
410	158	121.566
420	160	203.113
430	160	303.113
440	155	388.925
450	145	388.317
460	135	388.057
480	118	388.379
540	86	388.398
600	68	388.398
780	45	388.398
900	39	388.398

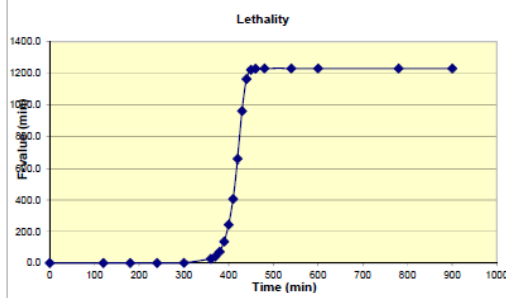
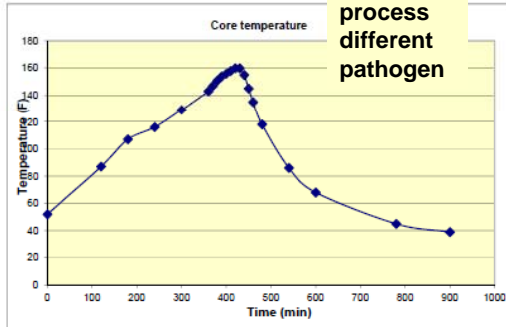
Note: lethality continues during cooling

PROCESS LETHALITY DETERMINATION

28-Sep-11
 Listeria Monocytogenes
 Ham of 9/23

User Must:
 1. Identify organism and product of concern
 2. Provide at least 20 time/temp data points

Same process different pathogen



T ref= 144 °F
 z = 10.8 °F
 D = 3.3 min

Log Reduction of Process
 172.97

Data Table		
Time (min)	Core Temp (°F)	F-value (min)
0	52	0.000
120	87	0.000
180	107	0.012
240	118	0.100
300	129	1.402
360	143	26.867
370	147	40.385
380	151	72.103
390	154	136.502
400	156	243.239
410	158	408.732
420	160	657.162
430	160	980.189
440	155	1163.881
450	145	1230.794
460	135	1229.189
480	118	1230.676
540	86	1230.793
600	68	1230.794
780	45	1230.794
900	39	1230.794

Another Web Site Based Lethality Calculator

- Mullan, W.M.A. (2007). Calculator for determining the F value of a thermal process. [On-line]. Available from: <http://www.dairyscience.info/index.php/thermal-processing/134-f-value-thermal-process.html>. Accessed: 13 July, 2012.

A calculator for determining the F or P value of a thermal process

Reference Temp, °C: 121.1 Start time, min: 2 Recalculate

Z-Value, °C: 10 End time, min: 26

F value of process: 0 Interval, min: 2 Upload CSV data file

Notes:
 Import your own data and vary Z and Tr to model F values.
 Use the "new data grid" and "start and end time" controls along with the "time interval" control to define your input requirements.
 The no. of log cycles that a designated microbial population has been reduced can be calculated by dividing the F value by the D value at Tr. A 12 log cycle reduction is required for spores of *Clostridium botulinum* in 'commercially sterile' low acid canned foods.
 If you find this calculator difficult to use try a simplified version where you replace the 'dummy data' with your own.

Time (Min)	Temp (C)	Lethal Rate
0	Edit 0	0
2	Edit 0	0
4	Edit 0	0
6	Edit 0	0
8	Edit 0	0
10	Edit 0	0
12	Edit 0	0
14	Edit 0	0
16	Edit 0	0
18	Edit 0	0
20	Edit 0	0
22	Edit 0	0
24	Edit 0	0
26	Edit 0	0

Another Web Site Based Lethality Calculator

◆ Combase Predictor

- ★ www.combase.cc
- ★ Requires registration
- ★ Limited temperature ranges, all in °C
- ★ Time in hours

The screenshot shows the ComBase Predictor web application interface. The page is titled "ComBase Predictor" and has a navigation menu with "Predictive Models", "Combase Predictor", "About Predictor", and "Predictor Help". The main content area is divided into three tabs: "Growth model", "Thermal inactivation model" (which is selected), and "Non thermal survival model".

The "Thermal inactivation model" tab contains several input fields and a "Predict" button. The "Temperature" section has radio buttons for "Static" and "Changing", with "Changing" selected. Below it is a text box for "Input your temperature profile in the textbox" containing a table of time and temperature values. The "Water Activity" section has radio buttons for "a_w" and "Aw", with "Aw" selected. The "Observation Duration" section has a "Time(h)" input field set to "30".

The "Predict" button is highlighted in red. To the right of the input fields is a graph showing "Predicted decrease (log CFU/g)" on the y-axis (ranging from -12 to 0) and "Time (h)" on the x-axis (ranging from 0 to 30). The graph shows a sharp initial drop in log CFU/g, followed by a slower, linear decrease.

Below the graph is a table of "Predictions" with columns for "Time(h)", "Temp.(C)", and "Conc.(Log10 cells/g)".

Time(h)	Temp.(C)	Conc.(Log10 cells/g)
0.00	54.50	0.00
0.30	54.51	-0.61
0.61	54.52	-1.44
0.91	54.53	-2.32
1.21	54.54	-3.20
1.52	54.55	-4.10
1.82	54.56	-4.99
2.12	54.57	-5.89
2.42	54.58	-6.80
2.73	54.59	-7.71
3.03	54.60	-8.62
3.33	54.61	-9.54

At the bottom of the page, there is a "Contact us" link, a "Disclaimer" link, and the copyright notice "©2012 Combase".

Food Research Institute,
University of Wisconsin- Madison,
July 2012

Process lethality spreadsheet

Instructions for Using the AMI Process Lethality Determination Spreadsheet

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Definitions

- Thermal Death Time: This is the time in minutes, necessary to kill a given number of organisms at a specified temperature.
- T ref: The reference temperature used when establishing the D- and z-values.
- D-Value: This indicates time in minutes at a constant temperature, that is necessary to destroy 90% or 1 log of the organism present at a given reference temperature. A D-value at one temperature, along with a z-value, is used to define the heat resistance of a microorganism.
- z-Value: This is the temperature increase required to reduce the thermal death time by a factor of 10. It is the number of degrees between a 10-fold change (or log cycle) in a microorganism's heat resistance. The z-value is considered a constant for a given microorganism strain in a given product.
- F-Value: This is the process lethality or the time in minutes, at a specific temperature required to destroy a certain number of viable cells.

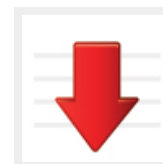
You must provide the following

- Identify microorganism and meat and/or poultry product of concern.
- Provide at least 20 time/core product temperatures that represents the products heating and cooling process.

Instructions:

1. By using the table that contains the lethality data from literature, we have selected the microorganism and product of concern. For example, let's say our organism of concern is *L. monocytogenes*, and our product is a hot dog. Identify the corresponding T ref (144 F), z-value (10.8 F), and D-value (3.3 min) provided in the table. These values should be obtained from your own companies challenge study data, from scientific literature, or other reliable sources. These values need to be relevant and appropriate for the type of product and the organism of concern. The table provides some example values from scientific literature that apply to certain products, but you need to justify your choice or provide more relevant values for your specific product and process.
2. Once the T ref (144 F), z-value (10.8 F), and D value (3.3 min) have been identified, enter them into the appropriate labeled cells below the table that contains the lethality data from literature.

Download spreadsheet



(/wp-

content/uploads/Process-Lethality-Spreadsheet-August-2010.xls)

3. The data table below these three values gives an example of what some time/temperature data points may look like. Time must be recorded in increasing minutes (0, 10 min, 20 min, 30 min) as each temperature value is recorded. The temperature must be the core product temperature that identifies the coolest spot in the product and the product should be in the coolest zone in the cooking chamber. It is suggested that at least 20 data points be entered into the data table. The values that you enter should be a time-temperature map of the product as it heats and cools.
4. Once the table has been completed, the F-value, or process lethality, will be calculated at each data point and a cumulative F-value will be given as the very last number in the right hand column of the data table (76.90 min). This number adds up the lethality values for each time interval and calculates an approximation of the area under the lethal rate curve. This value will be referred to as the “computed cumulative F value” or the “cumulative process lethality”. In the given example, the calculation results in an equivalent lethality at 144 F of 76.90 minutes. Clear the values in the first two columns and enter your own continuous process time and core product temperature (°F) in the appropriate columns.
5. After the data has been entered, a core temperature and a lethality curve are produced. The first graph shows a plot of the core product time/temperature relationship and the second graph shows a plot of the data converted to lethal rates or the cumulative F-value. In the example, because 144°F is the selected reference temperature, the area under the curve represents the total lethal effect of the process equivalent to 144 F. In this example, the lethality of the process is 76.90 minutes. This represents an “equivalent” time.
6. The total log reduction of the process is automatically determined by dividing the cumulative F-value (76.90) by the D-value (3.3) that was entered into the appropriate labeled cell. The resulting value equals the total log reduction of the process (23.30).
7. By using these estimates, you or a process authority should determine if the process meets regulatory requirements as safe. Additional documents, such as [Appendix A \(http://www.fsis.usda.gov/oa/fr/95033F-a.htm\)](http://www.fsis.usda.gov/oa/fr/95033F-a.htm), which discuss desired log reductions should also be considered when evaluating a lethality process.

Summary

This spreadsheet is to be used as a tool to determine if a specific cooking process has provided sufficient time and temperature to achieve a required log reduction for a given microorganism. If the appropriate log reduction is achieved for the process and organism of concern, the data and graphs provided in the spreadsheet may be used as a component of the HACCP validation materials. If the cooking process does not result in the appropriate log reduction, the cooking process needs to be re-evaluated and additional time and or temperature may need to be applied to the process.

Please direct all questions to the AMIF staff at 202-587-4200.

References

- Fain, Jr., A.R., J.E. Line, A.B. Moran, L.M. Martin, R.V. Lechowich, J.M. Carosella, and W.L. Brown. 1991. Lethality of heat to *Listeria monocytogenes* Scott A: D-value and z-value determinations in ground beef and turkey. *J. Food Prot.* 54(10):756-761.
- Food Safety and Inspection Service (FSIS). 1999. [Appendix A: Compliance guidelines for meeting lethality performance standards for certain meat and poultry products.](http://www.fsis.usda.gov/oa/fr/95033F-a.htm) (<http://www.fsis.usda.gov/oa/fr/95033F-a.htm>)
- Juneja, V.K. 2003. A comparative heat inactivation study of indigenous microflora in beef with that of *Listeria monocytogenes*, *Salmonella* serotypes and *Escherichia coli* O157:H7. *Letters Appl. Microbiol.* 37:292-298.
- Juneja, V.K. and B.S. Marmer. 1999. Lethality of heat to *Escherichia coli* O157: D- and z-value determinations in turkey, lamb and pork. *Food Research Intern.* 32(1):23-28.
- Line, J.E., A.R. Fain, Jr., A.B. Moran, L.M. Martin, R.V. Lechowich, J.M. Carosella, W.L. Brown. 1991. Lethality of heat to *Escherichia coli* O157:H7: D-value and Z-value determinations in ground beef. *J. Food Prot.* 54(10):762-766.
- Mazzotta, A.S. and D.E. Gombas. 2001. Heat resistance of an outbreak strain of *Listeria*

monocytogenes in hot dog batter. *J. Food Prot.* 64(3):321-324.

- Scott, J. and L. Weddig. 1998. Principles of Integrated Time-Temperature Processing (<http://www.amif.org/wp-content/uploads/Principles-of-Integrated-Time-Temperature-Processing.pdf>) *Proc. Meat Indus. Research Conf.* Philadelphia, PA.

Compatibility

The process lethality determination spreadsheet model is compatible with either Microsoft Excel version 5.0 (the version of Excel that is packaged with Microsoft Office 95) or Microsoft Excel 97 (the version that is packaged with Microsoft Office 97). Microsoft Excel version 5.0 and Microsoft Excel 97 will work regardless if the operating system is Windows 95, Windows 98 or Windows NT.



D-Value References

Compiled by M. E. Doyle

Food Research Institute, University of Wisconsin-Madison

Supplement to the FRI Webinar “Calculating Thermal Inactivation of Pathogens”

July 2012, updated Sept. 2012

1. Al-Holy M, Quinde Z, Guan D, Tang J, and Rasco B. Thermal inactivation of *Listeria innocua* in salmon (*Oncorhynchus keta*) caviar using conventional glass and novel aluminum thermal-Death-time tubes. *J Food Prot.* 2004; 67(2):383–386.
2. Al-Holy MA, Lin M, Abu-Ghoush MM, Al-Qadiri HM, and Rasco BA. Thermal resistance, survival and inactivation of *Enterobacter sakazakii* (*Cronobacter* spp.) in powdered and reconstituted infant formula. *J Food Safety.* 2009; 29(2):287–301.
3. Al Sakkaf A and Jones G. Thermal inactivation of *Campylobacter jejuni* in broth. *J Food Prot.* 2012; 75(6):1029–35.
4. Alvarez-Ordóñez A, Fernández ANA, Bernardo ANA, and López M. A comparative study of thermal and acid inactivation kinetics in fruit juices of *Salmonella enterica* serovar Typhimurium and *Salmonella enterica* serovar Senftenberg grown at acidic conditions. *Foodborne Path Dis.* 2009; 6(9):1147–1155.
5. Archer J, Jervis ET, Bird J, and Gaze JE. Heat resistance of *Salmonella weltevreden* in low-moisture environments. *J Food Prot.* 1998; 61(8):969–973.
6. Arroyo C, Condon S, and Pagan R. Thermobacteriological characterization of *Enterobacter sakazakii*. *Int J Food Microbiol.* 2009; 136(1):110–118.
7. Bacon RT, Ransom JR, Sofos JN, Kendall PA, Belk KE, and Smith GC. Thermal inactivation of susceptible and multiantimicrobial-resistant *Salmonella* strains grown in the absence or presence of glucose. *Appl Environ Microbiol.* 2003; 69(7):4123–4128.
8. Becker B, Schillinger U, Bohringer B, Untucht T, Izykowski N, and Franz C. Presence of bacilli in pasteurized packaged foods and determination of heat resistance of vegetative cells and spores of selected *Bacillus* isolates. *Archiv Lebensmittelhyg.* 2011; 62(6):205–211.
9. Benembarek PK and Huss HH. Heat-resistance of *Listeria monocytogenes* in vacuum-packaged pasteurized fish fillets. *Int J Food Microbiol.* 1993; 20(2):85–95.
10. Bermudez-Aguirre D and Corradini MG. Inactivation kinetics of *Salmonella* spp. under thermal and emerging treatments: a review. *Food Res Int.* 2012; 45(2):700–712.
11. Beuchat LR, Brackett RE, Hao DYY, and Conner DE. Growth and thermal inactivation of *Listeria monocytogenes* in cabbage and cabbage juice. *Can J Microbiol.* 1986; 32(10):791–795.
12. Beuchat LR and Mann DA. Inactivation of *Salmonella* on pecan nutmeats by hot air treatment and oil roasting. *J Food Prot.* 2011; 74(9):1441–1450.
13. Black DG, Ye XP, Harte F, and Davidson PM. Thermal inactivation of *Escherichia coli* O157:H7 when grown statically or continuously in a chemostat. *J Food Prot.* 2010; 73(11):2018–2024.
14. Bolton DJ, McMahon CM, Doherty AM, Sheridan JJ, McDowell DA, Blair LS, and Harrington D. Thermal inactivation of *Listeria monocytogenes* and *Yersinia enterocolitica* in minced beef under laboratory conditions and in sous-vide prepared minced and solid beef cooked in a commercial retort. *J Appl Microbiol.* 2000; 88(4):626–632.
15. Brackett RE, Schuman JD, Ball HR, and Scouten AJ. Thermal inactivation kinetics of *Salmonella* spp. within intact eggs heated using humidity-controlled air. *J Food Prot.* 2001; 64(7):934–938.
16. Bradshaw JG, Peeler JT, Corwin JJ, Hunt JM, Tierney JT, Larkin EP, and Twedt RM. Thermal resistance of *Listeria monocytogenes* in milk. *J Food Prot.* 1985; 48(9):743–745.
17. Brandl MT, Pan Z, Huynh S, Zhu YI, and McHugh TH. Reduction of *Salmonella enteritidis* population sizes on almond kernels with infrared heat. *J Food Prot.* 2008; 71(5):897–902.
18. Breidt F, Sandeep KP, and Arritt FM. Use of linear models for thermal processing of acidified foods. *Food Prot Trends.* 2010; 30(5):268–272.
19. Bremer PJ and Osborne CM. Thermal death times of *Listeria monocytogenes* in green shell mussels (*Perna canaliculus*) prepared for hot smoking. *J Food Prot.* 1995; 58(6):604–608.
20. Buduamoako E, Toora S, Walton C, Ablett RF, and Smith J. Thermal death times for *Listeria monocytogenes* in lobster meat. *J Food Prot.* 1992; 55(3):211–213.

21. Byrne B, Dunne G, and Bolton DJ. Thermal inactivation of *Bacillus cereus* and *Clostridium perfringens* vegetative cells and spores in pork luncheon roll. *Food Microbiol.* 2006; 23(8):803–808.
22. Carlson TR, Marks BP, Booren AM, Ryser ET, and Orta-Ramirez A. Effect of water activity on thermal inactivation of *Salmonella* in ground turkey. *J Food Sci.* 2005; 70(7):M363–M366.
23. Casadei MA, De Matos RE, Harrison ST, and Gaze JE. Heat resistance of *Listeria monocytogenes* in dairy products as affected by the growth medium. *J Appl Bacteriol.* 1998; 84(2):234–239.
24. Chang SS, Han AR, Reyes-De-Corcuera JI, Powers JR, and Kang DH. Evaluation of steam pasteurization in controlling *Salmonella* serotype Enteritidis on raw almond surfaces. *Lett Appl Microbiol.* 2010; 50(4):393–398.
25. Chantarapanont W, Slutsker L, Tauxe RV, and Beuchat LR. Factors influencing inactivation of *Salmonella enteritidis* in hard-cooked eggs. *J Food Prot.* 2000; 63(1):36–43.
26. Chhabra AT, Carter WH, Linton RH, and Cousin MA. A predictive model to determine the effects of pH, milkfat, and temperature on thermal inactivation of *Listeria monocytogenes*. *J Food Prot.* 1999; 62(10):1143–1149.
27. Couvert O, Gaillard S, Savy N, Mafart P, and Leguerinel I. Survival curves of heated bacterial spores: effect of environmental factors on Weibull parameters. *Int J Food Microbiol.* 2005; 101(1):73–81.
28. Daoust JY, Park CE, Szabo RA, Todd ECD, Emmons DB, and McKellar RC. Thermal inactivation of *Campylobacter* species, *Yersinia enterocolitica*, and hemorrhagic *Escherichia coli* O157:H7 in fluid milk. *J Dairy Sci.* 1988; 71(12):3230–3236.
29. De Jesus AJ and Whiting RC. Thermal inactivation, growth, and survival studies of *Listeria monocytogenes* strains belonging to three distinct genotypic lineages. *J Food Prot.* 2003; 66(9):1611–1617.
30. de Jong AEI, van Asselt ED, Zwietering MH, Nauta MJ, and de Jonge R. Extreme heat resistance of food borne pathogens *Campylobacter jejuni*, *Escherichia coli*, and *Salmonella* Typhimurium on chicken breast fillet during cooking. *Int J Microbiol.* 2012; (Epub 29 Jan):196841.
31. De Paula CMD, Mariot RF, and Tondo EC. Thermal inactivation of *Salmonella enteritidis* by boiling and frying egg methods. *J Food Safety.* 2005; 25(1):43–57.
32. Doherty AM, McMahon CMM, Sheridan JJ, Blair IS, McDowell DA, and Hegarty T. Thermal resistance of *Yersinia enterocolitica* and *Listeria monocytogenes* in meat and potato substrates. *J Food Safety.* 1998; 18(2):69–83.
33. Dorsa WJ, Marshall DL, Moody MW, and Hackney CR. Low-temperature growth and thermal inactivation of *Listeria monocytogenes* in precooked crawfish tail meat. *J Food Prot.* 1993; 56(2):106–109.
34. Doyle ME and Mazzotta AS. Review of studies on the thermal resistance of salmonellae. *J Food Prot.* 2000; 63(6):779–795.
35. Doyle ME, Mazzotta AS, Wang T, Wiseman DW, and Scott VN. Heat resistance of *Listeria monocytogenes*. *J Food Prot.* 2001; 64(3):410–429.
36. Edelson-Mammel SG and Buchanan RL. Thermal inactivation of *Enterobacter sakazakii* in rehydrated infant formula. *J Food Prot.* 2004; 67(1):60–63.
37. Elshenawy MA, Yousef AE, and Marth EH. Thermal inactivation and injury of *Listeria monocytogenes* in reconstituted nonfat dry milk. *Milchwissenschaft-Milk Sci Int.* 1989; 44(12):741–745.
38. Enache E, Mathusa, EC, Elliott PH, Black DG, Chen Y, Scott VN, and Schaffner DW. Thermal resistance parameters for Shiga toxin-producing *Escherichia coli* in apple juice. *J Food Prot.* 2011; 74(8):1231–1237.
39. Fain AR, Line JE, Moran AB, Martin LM, Lechowich RV, Carosella JM, and Brown WL. Lethality of heat to *Listeria monocytogenes* Scott A: D-value and Z-value determinations in ground beef and turkey. *J Food Prot.* 1991; 54(10):756–761.
40. Fairchild TM, Swartzel KR, and Foegeding PM. Inactivation kinetics of *Listeria innocua* in skim milk in a continuous-flow processing system. *J Food Sci.* 1994; 59(5):960–963.
41. Favier GI, Escudero ME, and De Guzman AMS. Thermal inactivation of *Yersinia enterocolitica* in liquid egg products. *J Food Safety.* 2008; 28(2):157–169.
42. Felicio MTS, Ramalheira R, Ferreira V, Brandao T, Silva J, Hogg TIM, and Teixeira P. Thermal inactivation of *Listeria monocytogenes* from Alheiras, traditional Portuguese sausage during cooking. *Food Control.* 2011; 22(12):1960–1964.
43. Feng G, Churey JJ, and Worobo RW. Thermal inactivation of *Salmonella* and *Escherichia coli* O157:H7 on alfalfa seeds. *J Food Prot.* 2007; 70(7):1698–1703.
44. Fernandez A, Lopez M, Bernardo A, Condon S, and Raso J. Modelling thermal inactivation of *Listeria monocytogenes* in sucrose solutions of various water activities. *Food Microbiol.* 2007; 24(4):372–379.
45. Gabriel AA. D values of composite *Salmonella enterica* serotypes typhimurium and enteritidis in Philippine native orange juice. *J Food Process Preserv.* 2007; 31(6):649–658.
46. Gabriel AA. Inactivation of *Escherichia coli* O157:H7 and spoilage yeasts in germicidal UV-C-irradiated and heat-treated clear apple juice. *Food Control.* 2012; 25(2):425–432.
47. Gabriel AA and Azanza MPV. D-72 degrees C values of *Salmonella typhimurium* in citrus juices: predictive efficacy of a model. *J Food Process Engin.* 2010; 33(3):506–518.
48. Gabriel AA, Barrios EB, and Azanza MPV. Modeling the thermal death of *Salmonella typhimurium* in citrus systems. *J Food Process Engin.* 2008; 31 (5):640–657.

49. Gabriel AA and Nakano H. Effects of culture conditions on the subsequent heat inactivation of *E. coli* O157:H7 in apple juice. *Food Control*. 2011; 22(8):1456–1460.
50. Gabriel AA and Nakano H. Inactivation of *Salmonella*, *E. coli* and *Listeria monocytogenes* in phosphate-buffered saline and apple juice by ultraviolet and heat treatments. *Food Control*. 2009; 20(4):443–446.
51. Gonzalez I, Lopez M, Fernandez A, and Bernardo A. Thermal inactivation of *Bacillus cereus* spores formed in media with different mineral contents. *Archiv Lebensmittelhyg*. 2008; 59(1):34–38.
52. Gonzalez I, Lopez M, Martinez S, Bernardo A, and Gonzalez J. Thermal inactivation of *Bacillus cereus* spores formed at different temperatures. *Int J Food Microbiol*. 1999; 51(1):81–84.
53. Gurtler JB, Marks HM, Jones DR, Bailey RR, and Bauer NE. Modeling the thermal inactivation kinetics of heat-resistant *Salmonella* Enteritidis and Oranienburg in 10 percent salted liquid egg yolk. *J Food Prot*. 2011; 74(6):882–892.
54. Gurtler JB, Rivera RB, Zhang HQ, and Sommers CH. Behavior of avirulent *Yersinia pestis* in liquid whole egg as affected by storage temperature, antimicrobials and thermal pasteurization. *J Food Safety*. 2010; 30(3):537–557.
55. Hajmeer MN, Tajkarimi M, Gomez EL, Lim N, O'Hara M, Riemann HP, and Cliver DO. Thermal death of bacterial pathogens in linguica smoking. *Food Control*. 2011; 22(5):668–672.
56. Harris LJ, Uesugi AR, Abd SJ, and McCarthy KL. Survival of *Salmonella enteritidis* PT 30 on inoculated almond kernels in hot water treatments. *Food Res Int*. 2012; 45(2):1093–1098.
57. He Y, Guo D, Yang J, Tortorello ML, and Zhang W. Survival and heat resistance of *Salmonella enterica* and *Escherichia coli* O157:H7 in peanut butter. *Appl Environ Microbiol*. 2011; 77(23):8434–8438.
58. Holsinger VH, Smith PW, Smith JL, and Palumbo SA. Thermal-destruction of *Listeria monocytogenes* in ice-cream mix. *J Food Prot*. 1992; 55(4):234–237.
59. Huang IPD, Yousef AE, Marth EH, and Matthews ME. Thermal inactivation of *Listeria monocytogenes* in chicken gravy. *J Food Prot*. 1992; 55(7):492–496.
60. Huang LH. Thermal inactivation of *Listeria monocytogenes* in ground beef under isothermal and dynamic temperature conditions. *J Food Engin*. 2009; 90(3):380–387.
61. Huang LH. Thermal resistance of *Listeria monocytogenes*, *Salmonella* Heidelberg, and *Escherichia coli* O157:H7 at elevated temperatures. *J Food Prot*. 2004; 67(8):1666–1670.
62. Ingham S. A comment on "Evaluation of additional Cooking Procedures to Achieve Lethality Microbiological Performance Standards for Large, Intact Meat Products" *J. Food Prot*. 74(10):1741–1745 (2011) (letter). *J Food Prot*. 2012; 75(4):629.
63. International Commission on Microbiological Specifications for Foods (ICMSF). *Microorganisms in Foods 5: Characteristics of Microbial Pathogens (Food Safety)*. 1996. Springer-Verlag New York.
64. Jagannath A, Tsuchido T, and Membre JM. Comparison of the thermal inactivation of *Bacillus subtilis* spores in foods using the modified Weibull and Bigelow equations. *Food Microbiol*. 2005; 22(2–3):233–239.
65. Jeong S, Marks BP, and Orta-Ramirez A. Thermal inactivation kinetics for *Salmonella enteritidis* PT30 on almonds subjected to moist-air convection heating. *J Food Prot*. 2009; 72(8):1602–1609.
66. Juneja VK. A comparative heat inactivation study of indigenous microflora in beef with that of *Listeria monocytogenes*, *Salmonella* serotypes and *Escherichia coli* O157:H7. *Lett Appl Microbiol*. 2003; 37(4):292–298.
67. Juneja VK. Heat resistance of enterohaemorrhagic *Escherichia coli* O157:H7 and *Salmonella*. *Bull Int Dairy Federation*. 2004(392):69–76.
68. Juneja VK. Thermal inactivation of *Salmonella* spp. in ground chicken breast or thigh meat. *Int J Food Sci Technol*. 2007; 42(12):1443–1448.
69. Juneja VK, Bari ML, Inatsu Y, Kawamoto S, and Friedman M. Thermal destruction of *Escherichia coli* O157:H7 in sous-vide cooked ground beef as affected by tea leaf and apple skin powders. *J Food Prot*. 2009; 72(4):860–865.
70. Juneja VK and Eblen BS. Heat inactivation of *Salmonella typhimurium* DT104 in beef as affected by fat content. *Lett Appl Microbiol*. 2000; 30(6):461–467.
71. Juneja VK and Eblen BS. Predictive thermal inactivation model for *Listeria monocytogenes* with temperature, pH, NaCl, and sodium pyrophosphate as controlling factors. *J Food Prot*. 1999; 62(9):986–993.
72. Juneja VK, Eblen BS, and Marks HM. Modeling non-linear survival curves to calculate thermal inactivation of *Salmonella* in poultry of different fat levels. *Int J Food Microbiol*. 2001; 70(1–2):37–51.
73. Juneja VK, Eblen BS, and Ransom GM. Thermal inactivation of *Salmonella* spp. in chicken broth, beef, pork, turkey, and chicken: determination of D- and Z-values. *J Food Sci*. 2001; 66(1):146–152.
74. Juneja VK, Huang L, and Yan X. Thermal inactivation of foodborne pathogens and the USDA pathogen modeling program. *J Therm Anal Calorimetry*. 2011; 106(1):191–198.
75. Juneja VK and Marks HM. Characterizing asymptotic D-Values for *Salmonella* spp. subjected to different heating rates in sous-vide cooked beef. *Innovative Food Sci Emerg Technolog*. 2003; 4(4):395–402.
76. Juneja VK, Marks HM, and Huang LH. Growth and heat resistance kinetic variation among various isolates of *Salmonella* and its application to risk assessment. *Risk Analysis*. 2003; 23(1):199–213.

77. Juneja VK and Marmer BS. Thermal inactivation of *Clostridium perfringens* vegetative cells in ground beef and turkey as affected by sodium pyrophosphate. *Food Microbiol.* 1998; 15(3):281–287.
78. Juneja VK, Novak JS, Eblen BS, and McClane BA. Heat resistance of *Clostridium perfringens* vegetative cells as affected by prior heat shock. *J Food Safety.* 2001; 21(2):127–139.
79. Juneja VK, Yadav AS, Hwang CA, Sheen S, Mukhopadhyay S, and Friedman M. Kinetics of thermal destruction of *Salmonella* in ground chicken containing trans-cinnamaldehyde and carvacrol. *J Food Prot.* 2012; 75(2):289–296.
80. Keller SE, Grasso EM, Halik LA, Fleischman GJ, Chirtel SJ, and Grove SF. Effect of growth on the thermal resistance and survival of *Salmonella* Tennessee and Oranienburg in peanut butter, measured by a new thin-layer thermal death time device. *J Food Prot.* 2012; 75(6):1125–1130.
81. Kennedy J, Blair IS, McDowell DA, and Bolton DJ. An investigation of the thermal inactivation of *Staphylococcus aureus* and the potential for increased thermotolerance as a result of chilled storage. *J Appl Bacteriol.* 2005; 99(5):1229–1235.
82. Kenney SJ and Beuchat LR. Survival, growth, and thermal resistance of *Listeria monocytogenes* in products containing peanut and chocolate. *J Food Prot.* 2004; 67(10):2205–2211.
83. Kim SH and Park JH. Thermal resistance and inactivation of *Enterobacter sakazakii* isolates during rehydration of powdered infant formula. *J Microbiol Biotechnol.* 2007; 17(2):364–368.
84. Kornacki JL and Marth EH. Thermal inactivation of *Staphylococcus aureus* in retentates from ultrafiltered milk. *J Food Prot.* 1989; 52(9):631–637.
85. Krapf T and Gantenbein-Demarchi C. Thermal inactivation of *Salmonella* spp. during conching. *Food Sci Technol.* 2010; 43(4):720–723.
86. Leguerinel I, Spegagne I, Couvert O, Gaillard S, and Mafart P. Validation of an overall model describing the effect of three environmental factors on the apparent D-value of *Bacillus cereus* spores. *Int J Food Microbiol.* 2005; 100(1–3):223–229.
87. Li X, Sheldon BW, and Ball HR. Thermal resistance of *Salmonella enterica* serotypes, *Listeria monocytogenes*, and *Staphylococcus aureus* in high solids liquid egg mixes. *J Food Prot.* 2005; 68(4):703–710.
88. Lihono MA, Mendonca AF, Dickson JS, and Dixon PM. Influence of sodium pyrophosphate on thermal inactivation of *Listeria monocytogenes* in pork slurry and ground pork. *Food Microbiol.* 2001; 18(3):269–276.
89. Line JE, Fain AR, Moran AB, Martin LM, Lechowich RV, Carosella JM, and Brown WL. Lethality of heat to *Escherichia coli* O157:H7: D-Value and Z-value determinations in ground-beef. *J Food Prot.* 1991; 54(10):762–766.
90. Lovett J, Bradshaw JG, and Peeler JT. Thermal inactivation of *Yersinia enterocolitica* in milk. *Appl Environ Microbiol.* 1982; 44(2):517–519.
91. Ma LI, Zhang G, Gerner-Smidt P, Mantripragada V, Ezeoke I, and Doyle MP. Thermal inactivation of *Salmonella* in peanut butter. *J Food Prot.* 2009; 72(8):1596–1601.
92. Mak PP, Ingram BH, and Ingham SC. Validation of apple cider pasteurization treatments against *Escherichia coli* O157:H7, *Salmonella*, and *Listeria monocytogenes*. *J Food Prot.* 2001; 64(11):1679–1689.
93. Manas P, Pagan R, Alvarez I, and Uson SC. Survival of *Salmonella senftenberg* 775 W to current liquid whole egg pasteurization treatments. *Food Microbiol.* 2003; 20(5):593–600.
94. Mazas M, Fernandez A, Alvarez A, Lopez M, and Bernardo A. Effects of phosphate and sodium and potassium chlorides on sporulation and heat resistance of *Bacillus cereus*. *J Food Safety.* 2009; 29(1):106–117.
95. Mazzotta AS. Heat resistance of *Listeria monocytogenes* in vegetables: evaluation of blanching processes. *J Food Prot.* 2001; 64(3):385–387.
96. Mazzotta AS. Thermal inactivation of stationary-phase and acid-adapted *Escherichia coli* O157:H7, *Salmonella*, and *Listeria monocytogenes* in fruit juices. *J Food Prot.* 2001; 64(3):315–320.
97. Mazzotta AS. Thermal inactivation of stationary-phase and salt-adapted *Listeria monocytogenes* during post-process pasteurization of surimi-based imitation crab meat. *J Food Prot.* 2001; 64(4):483–485.
98. McCormick K, Han IY, Acton JC, Sheldon BW, and Dawson PL. D- and Z-values for *Listeria monocytogenes* and *Salmonella typhimurium* in packaged low-fat ready-to-eat turkey bologna subjected to a surface pasteurization treatment. *Poult Sci.* 2003; 82(8):1337–1342.
99. Miles CA and Mackey BM. A mathematical analysis of microbial inactivation at linearly rising temperatures: calculation of the temperature rise needed to kill *Listeria monocytogenes* in different foods and methods for dynamic measurements of D-Value and Z-value. *J Appl Bacteriol.* 1994; 77(1):14–20.
100. Mogollon MA, Marks BP, Booren AM, Orta-Ramirez A, and Ryser ET. Effect of beef product physical structure on *Salmonella* thermal inactivation. *J Food Sci.* 2009; 74(7):M347–M351.
101. Monfort S, Sagarzazu N, Gayan E, Raso J, and Alvarez I. Heat resistance of *Listeria* species to liquid whole egg ultrapasteurization treatment. *J Food Engin.* 2012; 111(2):478–481.
102. Montville TJ, Dengrove R, De Siano T, Bonnet M, and Schaffner DW. Thermal resistance of spores from virulent strains of *Bacillus anthracis* and potential surrogates. *J Food Prot.* 2005; 68(11):2362–2366.
103. Moussa-Boudjemaa B, Gonzalez J, and Lopez M. Heat resistance of *Bacillus cereus* spores in carrot extract acidified with different acidulants. *Food Control.* 2006; 17(10):819–824.

104. Murphy RY, Beard BL, Martin EM, Keener AE, and Osaili T. Predicting process lethality of *Escherichia coli* O157:H7, *Salmonella*, and *Listeria monocytogenes* in ground, formulated, and formed beef/turkey links cooked in an air impingement oven. *Food Microbiol.* 2004; 21(5):493–499.
105. Murphy RY, Beard BL, Martin EM./Duncan LK, and Marcy JA. Comparative study of thermal inactivation of *Escherichia coli* O157:H7, *Salmonella*, and *Listeria monocytogenes* in ground pork. *J Food Sci.* 2004; 69(4):M97–M101.
106. Murphy RY, Duncan LK, Beard BL, and Driscoll KH. D and Z values of *Salmonella*, *Listeria innocua*, and *Listeria monocytogenes* in fully cooked poultry products. *J Food Sci.* 2003; 68(4):1443–1447.
107. Murphy RY, Duncan LK, Berrang ME, Marcy JA, and Wolfe RE. Thermal inactivation D- and Z-values of *Salmonella* and *Listeria innocua* in fully cooked and vacuum packaged chicken breast meat during postcook heat treatment. *Poult Sci.* 2002; 81(10):1578–1583.
108. Murphy RY, Duncan LK, Driscoll KH, and Marcy JA. Lethality of *Salmonella* and *Listeria innocua* in fully cooked chicken breast meat products during postcook in-package pasteurization. *J Food Prot.* 2003; 66(2):242–248.
109. Murphy RY, Duncan LK, Johnson ER, Davis MD, and Marcy JA. Thermal inactivation of *Salmonella senftenberg* and *Listeria innocua* in beef/turkey blended patties cooked via fryer and/or air convection oven. *J Food Sci.* 2002; 67(5):1879–1885.
110. Murphy RY, Duncan LK, Johnson ER, Davis MD, and Smith JN. Thermal inactivation D- and Z-values of *Salmonella* serotypes and *Listeria innocua* in chicken patties, chicken tenders, franks, beef patties, and blended beef and turkey patties. *J Food Prot.* 2002; 65(1):53–60.
111. Murphy RY, Johnson ER, Duncan LK, Davis MD, Johnson MG, and Marcy JA. Thermal inactivation of *Salmonella* spp. and *Listeria innocua* in the chicken breast patties processed in a pilot-scale air-convection oven. *J Food Sci.* 2001; 66(5):734–741.
112. Murphy RY, Johnson ER, Marks BP, Johnson MG, and Marcy JA. Thermal inactivation of *Salmonella senftenberg* and *Listeria innocua* in ground chicken breast patties processed in an air convection oven. *Poult Sci.* 2001; 80(4):515–521.
113. Murphy RY, Marks BP, Johnson ER, and Johnson MG. Inactivation of *Salmonella* and *Listeria* in ground chicken breast meat during thermal processing. *J Food Prot.* 1999; 62(9):980–985.
114. Murphy RY, Marks BP, Johnson ER, and Johnson MG. Thermal inactivation kinetics of *Salmonella* and *Listeria* in ground chicken breast meat and liquid medium. *J Food Sci.* 2000; 65(4):706–710.
115. Murphy RY, Martin EM, Duncan LK, Beard BL, and Marcy JA. Thermal process validation for *Escherichia coli* O157:H79, *Salmonella*, and *Listeria monocytogenes* in ground turkey and beef products. *J Food Prot.* 2004; 67(7):1394–1402.
116. Murphy RY, Osaili T, Duncan LK, and Marcy JA. Effect of sodium lactate on thermal inactivation of *Listeria monocytogenes* and *Salmonella* in ground chicken thigh and leg meat. *J Food Prot.* 2004; 67(7):1403–1407.
117. Murphy RY, Osaili T, Duncan LK, and Marcy JA. Thermal inactivation of *Salmonella* and *Listeria monocytogenes* in ground chicken thigh/leg meat and skin. *Poult Sci.* 2004; 83(7):1218–1225.
118. Nguyen HTT, Corry JEL, and Miles CA. Heat resistance and mechanism of heat inactivation in thermophilic campylobacters. *Appl Environ Microbiol.* 2006; 72(1):908–913.
119. Novak JS, Call J, Tomasula P, and Luchansky JB. An assessment of pasteurization treatment of water, media, and milk with respect to *Bacillus* spores. *J Food Prot.* 2005; 68(4):751–757.
120. O'Bryan CA, Crandall PG, Martin EM, Griffis CL, and Johnson MG. Heat resistance of *Salmonella* spp., *Listeria monocytogenes*, *Escherichia coli* O157:H7 and *Listeria innocua* M1, a potential surrogate for *Listeria monocytogenes*, in meat and poultry: a review. *J Food Sci.* 2006; 71(3):R23–R30.
121. Ollinger-Snyder P, El Gazzar F, Matthews ME, Marth EH, and Unklesbay N. Thermal-destruction of *Listeria monocytogenes* in ground pork prepared with and without soy hulls. *J Food Prot.* 1995; 58(5):573–576.
122. Onwuka UN, Akobundu ENT, and Iwe MO. Kinetics of inactivation of *Listeria monocytogenes*, *Clostridium perfringens*, *Escherichia coli* and *Salmonella* spp. in ohmic heated tomato juices. *J Pure Appl Microbiol.* 2008; 2(1):29–38.
123. Osaili T, Griffis CL, Martin EM, Beard BL, Keener A, and Marcy JA. Thermal Inactivation studies of *Escherichia coli* O157 : H7, *Salmonella*, and *Listeria monocytogenes* in ready-to-eat chicken-fried beef patties. *J Food Prot.* 2006; 69(5):1080–1086.
124. Osaili TM, Griffis CL, Martin EM, Beard BL, Keener AE, and Marcy JA. Thermal inactivation of *Escherichia coli* O157:H7, *Salmonella*, and *Listeria monocytogenes* in breaded pork patties. *J Food Sci.* 2007; 72(2):M56–M61.
125. Osaili TM, Shaker RR, Al-Haddaq MS, Al-Nabulsi AA, and Holley RA. Heat resistance of *Cronobacter* species (*Enterobacter sakazakii*) in milk and special feeding formula. *J Appl Bacteriol.* 2009; 107(3):928–935.
126. Oteiza JM, Giannuzzi L, and Califano AN. Thermal inactivation of *Escherichia coli* O157:H7 and *Escherichia coli* isolated from morcilla as affected by composition of the product. *Food Res Int.* 2003; 36(7):703–712.
127. Pagan R, Manas P, Alvarez I, and Sala FJ. Heat resistance in different heating media of *Listeria monocytogenes* ATCC 15313 grown at different temperatures. *J Food Safety.* 1998; 18(3):205–219.
128. Pagan R, Manas P, Raso J, and Trepal FJS. Heat resistance of *Yersinia enterocolitica* grown at different temperatures and heated in different media. *Int J Food Microbiol.* 1999; 47(1–2):59–66.

129. Park C, Stankiewicz Z, Daoust JY, and Emmons D. Thermal inactivation of *Yersinia* in fluid milk. *Can Inst Food Sci Technol J*. 1987; 20(5):323.
130. Pearce LE, Smythe BW, Crawford RA, Oakley E, Hathaway SC, and Shepherd JM. Pasteurization of milk: the heat inactivation kinetics of milk-borne dairy pathogens under commercial-type conditions of turbulent flow. *J Dairy Sci*. 2012; 95(1):20–35.
131. Porto-Fett ACS, Juneja VK, Tamplin ML, and Luchansky JB. Validation of cooking times and temperatures for thermal inactivation of *Yersinia pestis* strains KIM5 and CDC-A1122 in irradiated ground beef. *J Food Prot*. 2009; 72(3):564–571.
132. Pucciarelli AB and Benassi FO. Inactivation of *Salmonella enteritidis* on raw poultry using microwave heating. *Braz Arch Biol Technol*. 2005; 48(6):939–946.
133. Rahman MS, Guizani N, and Al-Ruzeiki MH. D- and Z-values of microflora in tuna mince during moist- and dry-heating. *Food Sci Technol*. 2004; 37(1):93–98.
134. Rajkowski KT. Thermal inactivation of *Escherichia coli* O157:H7 and *Salmonella* on catfish and tilapia. *Food Microbiol*. 2012; 30(2):427–431.
135. Rustia AS and Azanza MPV. Heat resistance characteristics of *Salmonella enteritidis* in liquid quail egg. *Food Sci Technol Res*. 2005; 11(2):151–156.
136. Sallami L, Marcotte M, Naim F, Ouattara B, Leblanc C, and Saucier L. Heat inactivation of *Listeria monocytogenes* and *Salmonella enterica* serovar Typhi in a typical bologna matrix during an industrial cooking-cooling cycle. *J Food Prot*. 2006; 69(12):3025–3030.
137. Schoeni JL, Brunner K, and Doyle MP. Rates of thermal inactivation of *Listeria monocytogenes* in beef and fermented beaker sausage. *J Food Prot*. 1991; 54(5):334–337.
138. Schultze KK, Linton RH, Cousin MA, Luchansky JB, and Tamplin ML. Effect of preinoculation growth media and fat levels on thermal inactivation of a serotype 4b strain of *Listeria monocytogenes* in frankfurter slurries. *Food Microbiol*. 2007; 24(4):352–361.
139. Sevilla KP and Gabriel AA. D values of *Escherichia coli* in tilapia meat. *J Muscle Foods*. 2010; 21(2):167–176.
140. Sharma M, Adler BB, Harrison MD, and Beuchat LR. Thermal tolerance of acid-adapted and unadapted *Salmonella*, *Escherichia coli* O157:H7, and *Listeria monocytogenes* in cantaloupe juice and watermelon juice. *Lett Appl Microbiol*. 2005; 41(6):448–453.
141. Silva FVM and Gibbs PA. Thermal pasteurization requirements for the inactivation of *Salmonella* in foods. *Food Res Int*. 2012; 45(2):695–699.
142. Smith SE, Maurer JL, Orta-Ramirez A, Ryser ET, and Smith DM. Thermal inactivation of *Salmonella* spp., *Salmonella typhimurium* DT104, and *Escherichia coli* O157:H7 in ground beef. *J Food Sci*. 2001; 66(8):1164–1168.
143. Solomon EB, Huang LH, Sites JE, and Annous BA. Thermal inactivation of *Salmonella* on cantaloupes using hot water. *J Food Sci*. 2006; 71(2):M25–M30.
144. Sorqvist S. Heat resistance in liquids of *Enterococcus* spp., *Listeria* spp., *Escherichia coli*, *Yersinia enterocolitica*, *Salmonella* spp. and *Campylobacter* spp. *Acta Vet Scand*. 2003; 44(1–2):1–19.
145. Sorqvist S. Heat resistance of different serovars of *Listeria monocytogenes*. *J Appl Bacteriol*. 1994; 76(4):383–388.
146. Stopforth JD, Suhaimi R, Kottapalli B, Hill WE, and Samadpour M. Thermal inactivation D- and Z-values of multidrug-resistant and non-multidrug-resistant *Salmonella* serotypes and survival in ground beef exposed to consumer-style cooking. *J Food Prot*. 2008; 71(3):509–515.
147. Tuntivanich V, Orta-Ramirez A, Marks BP, Ryser ET, and Booren AM. Thermal inactivation of *Salmonella* in whole muscle and ground turkey breast. *J Food Prot*. 2008; 71(12):2548–2551.
148. Van Asselt ED and Zwietering MH. A systematic approach to determine global thermal inactivation parameters for various food pathogens. *Int J Food Microbiol*. 2006; 107(1):73–82.
149. Veeramuthu GJ, Price JF, Davis CE, Booren AM, and Smith DM. Thermal inactivation of *Escherichia coli* O157:H7, *Salmonella senftenberg* and enzymes with potential as time-temperature indicators in ground turkey thigh meat. *J Food Prot*. 1998; 61(2):171–175.
150. Velasquez A, Breslin TJ, Marks BP, Orta-Ramirez A, Hall NO, Booren AM, and Ryser ET. Enhanced thermal resistance of *Salmonella* in marinated whole muscle compared with ground pork. *J Food Prot*. 2010; 73(2):372–375.
151. Weiss A and Hammes WP. Efficacy of heat treatment in the reduction of salmonellae and *Escherichia coli* O157:H- on alfalfa, mung bean and radish seeds used for sprout production. *Eur Food Res Technol*. 2005; 221(1–2):187–191.
152. Wescott GG, Fairchild TM, and Foegeding PM. *Bacillus cereus* and *Bacillus stearothermophilus* spore inactivation in batch and continuous-flow systems. *J Food Sci*. 1995; 60(3):446–450.
153. Xu S, Labuza TP, and Diez-Gonzalez F. Thermal inactivation of *Bacillus anthracis* spores in cow's milk. *Appl Environ Microbiol*. 2006; 72(6):4479–4483.
154. Yuk HG, Geveke DJ, Zhang HQ, and Jin TZ. Comparison of aluminum thermal-death-time disks with a pilot-scale pasteurizer on the thermal inactivation of *Escherichia coli* K12 in apple cider. *Food Control*. 2009; 20(11):1053–1057.